Genetic Variation Among Morphological Forms of Pelagic Sargassum and Associated Hydroids

Variación Genética Entre las Formas Morfológicas de Sargassum Pelágico y Hidrozoos Asociado

Variation Génétique Entre les Formes Morphologiques de Sargassum Pélagique et Hydroids Associé

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## EXTENDED ABSTRACT

Pelagic Sargassum is comprised of two species distinguished by stem characteristics. S. fluitans has thorny stems and S. natans has smooth stems (Parr 1939). Each species is further divided into multiple morphological forms, although S. fluitans III Parr and S. natans I Parr historically dominated abundance by 90% or more (Parr 1939). Until recently, pelagic Sargassum observations indicated abundance maxima in the Sargasso Sea, at the center of the North Atlantic gyre, and the Gulf of Mexico (Parr 1939, Gower and King 2011, Hu et al. 2016). The Caribbean region, Brazil, and the west African coast have experienced episodic inundations of pelagic Sargassum since 2011. Coincident with the recent abundance increase in the Caribbean, there has been a shift in pelagic Sargassum diversity. In the western tropical Atlantic, eastern Caribbean and Antilles Current (north of Puerto Rico) during late 2014, Schell et al. (2015) documented mixed assemblages of pelagic Sargassum overwhelmingly dominated by the previously rare S. natans VIII Parr. However, with broad leaves and bladders often lacking spines, S. natans VIII Parr was mistaken for S. fluitans III Parr in some initial reports from the region. To compound the confusion associated with identification, Aglaophenia latecarinata, a hydroid epibiont that was historically exclusive to and dominant on S. fluitans III Parr (Calder 1995), was also dominant on S. natans VIII Parr.

In an attempt to validate morphological species delineations in pelagic Sargassum and test the hypothesis that the recent influx of S. natans VIII Parr originated from an alternate source region, we collected Sargassum and hydroid specimens from the Sargasso Sea, western tropical Atlantic and eastern Caribbean. During four separate Sea Education Association (Woods Hole, MA) cruises spanning from April 2015 through May 2016, we used both a dip net and neuston net to collect 44 pelagic Sargassum and 53 associated hydroid samples. We aimed to collect approximately 10 replicates at each sampling site. Sargassum and hydroid collections were not necessarily coincident due to distinct distribution patterns of the now three common forms of pelagic Sargassum. S. natans VIII Parr and associated hydroids were collected from the western tropical Atlantic during late 2015. S. fluitans III Parr samples were collected during late 2015 and early 2016 from the eastern Caribbean. Hydroids associated with S. fluitans III Parr were collected from the Antilles Current during April 2016. Additional hydroids were isolated in May 2016 from S. natans VIII Parr and S. fluitans III Parr collected simultaneously from the northern Gulf Stream. S. natans I Parr was collected from the Sargasso Sea in April and May 2015. Pelagic Sargassum abundance was extremely low in the Sargasso Sea during spring 2016, hence the lack of additional S. natans I Parr samples from this period. Sargassum was dried in silica gel and hydroid colonies were preserved in 95% ethanol prior to DNA extraction. We amplified DNA segments from three target genes for Sargassum and two target genes for the hydroid. For Sargassum, we sequenced the mitochondrial cytochrome oxydase 3 subunit (cox3), the Ribulose bisphosphate carboxylase large chain + spacer + partial Ribulose bisphosphate carboxylase small chain (rbcL-S), and the mitochondrial intergenic spacer region (mtsp) between the mitochondrial 23S gene and transcription ribonucleic acid Valine (tRNA-Val). For hydroids, we sequenced mitochondrial 16S rDNA and cytochrome c oxidase I (COI) markers. After assembly and alignment of bi-directional sequences, we created neighbor-joining trees and haplotype networks to visualize the relationships between pelagic Sargassum forms as wells as hydroids collected from each of the pelagic Sargassum substrates.

Sequences were identical across samples of individual pelagic Sargassum forms. S. fluitans III Parr sequences for all three markers were distinct from those of the S. natans-complex (Figure 1). There were ten polymorphic loci in the mtsp

region as well as two polymorphic loci and nine base pair insertion in rbcL-S region. With the present set of genetic markers, we could not distinguish between S. natans I Parr and S. natans VIII Parr despite clear morphological differences. Interestingly, we came across a few samples of what we provisionally identified as S. natans II Parr; these samples were genetically distinct from both the S. natanscomplex and S. fluitans III Parr (Figure 1). A. latecarinata sequences clustered into two distinct groups according to Sargassum substrate, even for samples collected from cooccurring S. natans VIII Parr and S. fluitans III Parr in the northern Gulf Stream (Figure 2). In the case of the mitochondrial 16S rRNA encoding region, we identified four distinct haplotypes with a maximum of a three base pair difference between sequences. Four distinct haplotypes were identified among sequences from the mitochondrial CO1 region, including one haplotype that represented a single hydroid sample collected from S. natans II Parr.

These preliminary genetic results support the morphological identification of S. natans VIII Parr by Schell et al. (2015) and the physical evidence for an alternate pelagic Sargassum source region associated with the recent inundations of S. natans VIII Parr (Gower et al. 2013; Franks et al. 2016). S. natans VIII Parr is genetically distinct from S. fluitans III Parr, despite superficial morphological similarities, and genetically indistinct, using the selected marker genes, from S. natans I Parr. Moreover, the associated hydroids were genetically distinct between Sargassum substrates. This result was anticipated, as A. latecarinata produces a planula larva that settles quickly and is dependent upon a substrate for dispersal. Together, these complementary genetic results reinforce the previous morphology-driven assertion that recent inundation events introduced a heretofore rare form of pelagic Sargassum to the Caribbean region that likely originated from the



**Figure 1.** Consensus Neighbor Joining Tree of the concatenated rbcL-S and mtsp loci. Bootstrap support is indicated at nodes, with those less than 95% supported collapsed. Number of replicates of each sequence shown in parentheses. Branch length is unconstrained and corresponds to genetic distance at a scale of 0.002. Outgroup sequences obtained from GenBank using NCBI Blast. Using morphological methods, *Sargassum* sp. has been provisionally identified as *S. natans* II Parr.

equatorial North Atlantic. This shift in regional Sargassum abundance and diversity not only has implications for coastal ecosystems and economies experiencing inundations, but also for pelagic populations that utilize Sargassum as a nursery or foraging habitat. A deeper examination of genetic as well as species and functional diversity among the S. natans VIII Parr associated community of organisms is warranted.

KEYWORDS: Sargassum, A glaophenia latecarinata, hydroid, population genetics, Caribbean

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**Figure 2.** Haplotype networks of the mt16S (top) and CO1 (bottom) genes in *A. latecarinata*. Circles and associated values depict number of samples within each distinct haplotype. Unbroken lines connecting circles represent a single nucleotide difference. Nucleotide changes are noted below each connecting line. Sequences from hydroids found on *S. natans* VIII Parr are in green, *S. fluitans* III Parr are in purple, and *S. natans* II Parr are in pink.